

**Title:** Comparative Review of Large Animal Models for Suitability of Proximal Aortic Endovascular Repair

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13

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1 **ABSTRACT.**

2

3 The advent of thoracic endovascular aortic repair (TEVAR) heralds a paradigm shift in treating descending  
4 aortopathies and is viewed as a potential option for ascending aortic dissection (AD) repair too. Currently,  
5 TEVAR's usage for ascending aortopathies remains limited. An urgent requirement exists to choose appropriate  
6 animal models to better contemporary understanding of endovascularly treating ascending ADs, also known as  
7 Stanford Type-A ADs. This narrative review provides a current literature summary on this topic, including the  
8 gross anatomical differences between adult porcine, ovine and bovine species versus that of their human  
9 counterparts, as well as specific valvular and coronary vasculature measurement variances. To achieve this,  
10 an electronic search of Cochrane Library, PubMed and Ovid Medline databases from January 1965 to June  
11 2020 was performed, limited to articles published in English. A total of twenty-three research papers were  
12 included in constructing this review. Our findings showed that whilst macroscopic anatomy remains grossly  
13 similar, differences in valvular leaflet shape are present, with porcine and ovine models possessing anatomic  
14 characteristics that are comparable to their human counterparts. Research into inter-species ascending aortic  
15 anatomy has not been extensively performed, highlighting a literature gap. Conversely, multiple morphological  
16 studies have highlighted that porcine coronary vasculature is closely resemblant to that of humans. In  
17 conclusion, both porcine and ovine species are suitable as appropriate animal models in examining the  
18 feasibility of endovascular stent-grafts for ascending ADs. However, given the similarities in coronary and aortic  
19 valve anatomy to their human analogues, porcine models are better suited for this purpose.

20

21 **Key Words:** Aortic dissection; Endovascular; Ascending aorta; Animal models (Source: MeSH-NLM).

22

## 1 INTRODUCTION.

2 The usage of non-human tissue in cardiothoracic medical research has markedly risen over the last half a  
3 century, as a solution to both the ethical dilemmas posed by using, as well as the lack of readily available  
4 human tissue for creating experimental clinical models.<sup>1</sup> One example of research involving such animal  
5 models is seen in better understanding treatment outcomes for acute aortic dissections (AD), a life-  
6 threatening pathology that carries significant mortality rates of over 70% within one week of onset when left  
7 untreated.<sup>2,3</sup> Several classifications of ADs currently exist, but arguably perhaps, one of the most commonly  
8 used is the Stanford classification system. This system categorizes dissections on the basis of site of  
9 intimomedial tear as either Type-A, defined as any AD involving the ascending aorta or Type-B, which are  
10 ADs not involving the ascending aorta (NB. This review focuses primarily on Type-A ADs).<sup>4</sup>

11

12 With few exceptions, the management of acute Type-A ADs is touted as a surgical emergency.<sup>5,6</sup> Given the  
13 aforementioned high rates of mortality otherwise, there are few reasons for not following through with  
14 operative treatment of Type-A ADs, with the main cited reasons being presence of significant medical  
15 comorbidities that impact survival to one year or less, as with very advanced age and frailty, advanced  
16 malignancies or patient/family wishes.<sup>7</sup> The surgical intervention for Type-A ADs has seen a marked evolution  
17 over the years, due to the intertwined combination of technological improvements in equipment, as well as a  
18 better understanding of its natural history. At present, open surgical repair (OSR) remains the gold standard of  
19 care for this otherwise catastrophic condition.<sup>4,8</sup> However, the advent of thoracic endovascular aortic repair  
20 (TEVAR) has heralded a paradigm shift in treatment options for aortic disease involving the descending aorta,  
21 and as such has been viewed as a potential option for ascending aortic repair, and consequently Type-A AD  
22 surgical repair as well.<sup>9</sup> As a result, selected patients who would otherwise be ineligible for OSR as  
23 aforementioned, which typically comprise up to 20% of all individuals, would benefit from having the  
24 opportunity of still receiving life-saving treatment in the form of minimally invasive endovascular techniques.<sup>10</sup>

25

26 There are various types of endovascular therapies currently viewed as a potential solution in treating Type-A  
27 ADs, including branched stent-grafts and valve-carrying conduits.<sup>10</sup> However, the employment of these novel  
28 therapeutic procedures within a clinical setting remains limited, with isolated case-reports and case-series  
29 providing the bulk of currently available literature on patient outcomes. Consequently, there exists an urgent  
30 requirement to choose appropriate animal models in order to better our understanding of endovascularly  
31 treating Type-A ADs.

32

33 While there is a widespread amount of published research on the variances of cardiothoracic anatomy in non-  
34 human species, there exists no literature review synthesizing this information, highlighting the accelerated  
35 need for one to be formulated. Consequently, this review article aims to combat this issue by providing a  
36 summary of currently available information on this topic, with a particular focus on determining which animal  
37 model amongst those of adult porcine, ovine or bovine species would be ideal for research pertaining to  
38 endovascularly treating Type-A ADs, relevant to the practising surgeon. Three broad sections shall be  
39 covered, beginning with a discussion of the macroscopic anatomical differences amongst humans, porcines,  
40 ovines and bovines. The review shall then focus on specific aspects of cardiothoracic anatomy, explicating in  
41 particular the valvular, aortic and coronary vasculature differences. Finally, the suitability of which animal

- 1 would be best for being used as clinical experimental models, from a strictly anatomical standpoint for
- 2 bettering our understanding of Type-A AD treatment shall then be explored.
- 3

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## 1 **METHODS**

2 To appropriately answer the aforementioned questions on this topic, two main databases were utilized. These  
3 included:

- 4 a) Ovid Medline
- 5 b) PubMed

6

7 Within Ovid Medline, since the term 'Type A aortic dissection' is quite well known within medical literature (as  
8 opposed to its verbatim analogue 'Stanford Type A aortic dissection'), the search string was commenced by  
9 initially mapping the keyword 'Endovascular' with the MeSH term 'Type A aortic dissection'. This was followed  
10 by using the Boolean operator 'AND'. The keyword 'models' was then utilized, and finally, the Boolean  
11 operator 'AND' was employed to combine all search strings. A total of 12 results were obtained from Ovid  
12 Medline. For the sake of this review, search results were limited to the English language. Secondly, within  
13 PubMed, an advanced search was conducted using the search terms 'endovascular', 'aortic dissection' and  
14 'animal model'. This result in a total of 26 results, which were then analyzed in conjunction with previous  
15 results obtained through Ovid Medline. A flow-chart of our search strategy and study selection is detailed  
16 below.

17

18 Finally, to obtain a better pictorial representation of the cardiothoracic anatomical variations between porcine,  
19 ovine and bovine models, images from the University of Minnesota Atlas of Human Cardiac Anatomy were  
20 used with permission.

## 1 RESULTS

### 2 **Anatomical considerations for endovascular therapy of Type-A dissections amongst** 3 **humans**

4 In spite of the advantages the utilisation of thoracic endovascular aortic repair (TEVAR) affords, including  
5 eliminating the need for perioperative cardiopulmonary bypass and the requirement for a major operative  
6 incision such as a sternotomy, there exist certain limitations that prevent its routine employment in currently  
7 treating Type-A ADs.<sup>4,11-13</sup> Given the paucity of large-scale trials documenting its efficacy however, as well as  
8 long-term follow-up of patients who receive this modality of treatment, there exists a literature gap in  
9 describing the specific limitations of endovascular therapy for ascending aortic pathologies. That being said,  
10 the anatomical constraints of this novel therapy have received particular scrutinization, and shall now be  
11 explored further.

12  
13 One of the major challenges in successfully treating Type-A ADs with currently available stent-grafts lies in the  
14 need to insert a straight device into a curved structure i.e. the aortic arch, which poses a high risk of  
15 developing an endoleak. In an attempt to simplify landmarks within the complex anatomy of the aortic arch,  
16 the Ishimaru classification is commonly used to categorize thoracic aortic 'zones' for stent-grafts.<sup>14</sup>

17  
18 Utilising Ishimaru's zone classifications, it is essential to ensure a 'safe' distance between the proximal and  
19 distal landing zones, to facilitate successful deployment of the stent-graft, as well as to avoid catastrophic  
20 aortic rupture.<sup>3,15,16</sup> However, this measurement remains dependent on both the characteristics of the chosen  
21 stent-graft, as well as the technical expertise of the operating doctor. Consequently, although there exists  
22 some variation in what constitutes a 'safe' distance, a proposed criterion has been a length of at least 20  
23 millimetres between the two landing zones, to avoid aortic rupture during graft deployment.<sup>16</sup>

24  
25 Secondly, problems are also created by the entry dissection tear occurring proximally within Zone 0 as  
26 illustrated in **Figure 1**, specifically proximal to the sinotubular junction. A tear occurring within this region  
27 would fail to allow endograft deployment in a manner that would allow coronary blood flow to be maintained.<sup>15</sup>  
28 Occlusion of the coronary ostia by closed ends of the stent-graft would cause ischaemia of the myocardium,  
29 resulting in potential irreversible damage.<sup>17,18</sup> Additionally, those with Type-A ADs extending into the aortic  
30 valve would not be suitable for endovascular treatment with conventional stent-grafts, a situation typically  
31 seen in between 10-20% of patients.<sup>15</sup> This is because at deployment the tip of the device must cross the  
32 aortic valve, which could eventuate in possible ventricular perforation. Although this would pose a barrier to  
33 treatment with currently available stent-grafts, given that they possess a distal cone that prevents their  
34 deployment too close to the aortic valve, a proposed method to combat this has been suggested in the form of  
35 novel 'valve-carrying conduits'.

36  
37 Thirdly, variations with the anatomy of the normal aorta may interfere with a wholly endovascular modality of  
38 treatment for Type-A ADs. For instance, in those who have received prior coronary artery bypass surgery, the  
39 presence of coronary grafts arising directly from the ascending aorta would present an increased risk of  
40 myocardial ischaemia during endograft deployment.<sup>15,16</sup>



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Based on these caveats, it is evident that the anatomy of the ascending aorta, aortic valve and coronary vasculature are of particular significance in determining an appropriate animal model for Type-A dissection research, which shall be addressed in the following sub-section.

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## 1 **Introduction and general cardiac anatomy**

2 Similar to those of humans, the holistic cardiac anatomy of large mammals is analogous; four cardiac valves  
3 are present with similar structures comparable to most quadruped mammals. Whilst human hearts can appear  
4 in a variety of shapes, including elliptical, trapezoidal, and 'valentine', porcine species tend to be valentine  
5 shaped, whilst the ovine heart varies from valentine to conical in shape, as illustrated in **Figure 2.**<sup>19</sup>

6

7 With respect to the hearts of porcine and ovine species, the distance between the posteroinferior base to  
8 apex, left lateral base to apex and the length of coronary sinuses are all significantly greater in their human  
9 counterparts. As expected therefore, in conjunction with its comparatively larger size, the average human  
10 heart maintains a larger organ to body weight ratio to that of both porcine and ovine species. A similar  
11 scenario is visible in that of bovines, which possesses a nearly identical organ to body weight ratio of the  
12 ovine species.<sup>19</sup>

13

## 14 **Valvular anatomy**

15 Whilst the general cardiac anatomy of different hearts remains roughly similar, variations in the four valves  
16 exist that distinguish porcine, ovine, bovine and human species, in spite of certain structural similarities.  
17 Illustrated in table 1, average aortic valve annulus diameters for humans are identical to that of their porcine  
18 counterparts, with the ovine species possessing a slightly narrower annulus on average. Conversely, bovine  
19 diameters are nearly 40% greater than their human counterparts, possibly accounted for due to the increased  
20 cardiac output within this species.<sup>20</sup>

21

22 Additionally, humans have much less muscular attachment surrounding the aortic valve compared to animal  
23 hearts, an indication of their reduced cardiac output.<sup>20</sup> On a similar note, the human aortic valve at the level of  
24 the annulus possesses muscular attachment along 43% of its circumference, compared to respective figures  
25 of 56%, 60%, and 57% in porcine, bovine, and ovine valves.<sup>20,21</sup> Additionally, a greater amount of myocardial  
26 tissue support is also present at the aortic valve's right and left coronary cusp bases, distinguishing all three of  
27 ovine, bovine, and porcine valves from the human aortic valve. It should be noted that in clinical trials  
28 involving sub-coronary transplantation, this increased muscle mass has been shown to result in aortic valvular  
29 stenosis.<sup>20</sup>

30

31 Differences in aortic valve leaflet shape and structure are also present, with only porcine valve leaflet depths  
32 being comparable to their human analogues, although specimen analysis visualized more inter-species  
33 variation between individual leaflets in the former.<sup>20</sup> Variations in leaflet thickness are particularly important to  
34 make note of, as thin and fragile leaflets such as those seen in ovine species may not be structurally strong  
35 enough to support heavy pressure loads during clinical usage for long periods of time.

36

## 37 **Aortic anatomy**

38 Unlike the aforementioned aspects of valvular anatomy, research into specifically the ascending aortic  
39 differences between human and non-human species has not been extensively performed, highlighting a  
40 current literature gap within this area. However, morphometric studies to determine the structural

1 characteristics of the largest artery in mammals have been documented. Primarily, compared to the human  
2 heart, the porcine species has only 2 head branches that originate from the aortic arch.

3  
4 Dimensionally, the diameter of the proximal aorta amongst porcine species at its largest part is about 21%  
5 lesser than that of their human analogues. What is noteworthy is that unlike their human counterparts, which  
6 exhibit a gradual diameter decrease in tapering fashion, there is a sharp decrease in aortic diameter from the  
7 descending thoracic aorta to the abdominal aorta within porcine, with exact values having been documented  
8 in Table 2. Conversely, whilst research on the aortic anatomy of ovine species is scarce, it is known that the  
9 ascending aorta itself, whilst maintaining a similar aortic diameter to that of their human counterparts after  
10 accounting for the changes in organ to body weight ratio, is relatively short, of which the implications shall be  
11 discussed in the next section.<sup>27</sup> There is also a marked decrease in the number of elastic lamellae within  
12 ovine aorta, greatly reducing its mobility as well.<sup>27</sup>

13  
14 Finally, of the three non-human species described in this paper, perhaps the most review has been done on  
15 bovine ascending aortic anatomy, with the 'bovine aortic arch' having been described as the single most  
16 common congenital aortic anatomic variant within humans as well. Whilst this term itself is a misnomer, it is  
17 used to supposedly refer to the variant within bovine species, which is characterised by a common single  
18 brachiocephalic trunk trifurcating into bilateral subclavian vessels and a single bicarotid trunk, as opposed to  
19 the more common human aortic arch which splits into a single brachiocephalic trunk, the left common carotid  
20 and the left subclavian arteries.<sup>28,29</sup>

21  
22 Similar to their ovine counterparts, little to no research has been done explicating the dimensional differences  
23 in aortic root diameter between bovines and humans, elucidating the need for further research in this area.

### 24 25 **Coronary anatomy**

26 The suitability of porcine species usage as an animal model in coronary arterial disease is well established,  
27 with multiple morphological studies highlighting that porcine coronary vasculature is closely resemblant to that  
28 of man.<sup>33</sup> In pigs, both coronary arteries arise from the aortic sinuses below the supra-avalvular ridge, as is seen  
29 in human species, with one study highlighting that all tested porcine models showed right coronary artery  
30 (RCA) dominance (humans typically exhibit RCA dominance of anywhere between 75 to 85%, depending on  
31 the chosen study analysed).<sup>34</sup> As with their human counterparts however, certain inter-species variants are  
32 present, and should be kept in mind whilst choosing a porcine animal model.<sup>34,35</sup>

33  
34 With regards to the coronary arterial system, in contrast to their porcine and human analogues, ovine species  
35 primarily have a left coronary type circulation; ergo, the majority of the myocardium receives its blood supply  
36 via branches of the left coronary artery.<sup>36</sup> However, given that ovines do not possess an extensive coronary  
37 collateral network, it may be still suitable to utilise their models for research. More specifically, although there  
38 exists considerable literature that is descriptive of specific aspects of ovine cardiac anatomy, little to no  
39 comparative research has been performed to elucidate the differences between ovine and human heart  
40 models, highlighting a significant literature gap in this area.<sup>36</sup>

1 The coronary vasculature of bovine species has also been studied and documented. In all examined animals,  
2 the coronary ostia were located beneath the sinotubular junction, as seen within their human counterparts.<sup>37</sup>  
3 The dimensions of coronary ostia are listed in Table 3, but it is important note that ovines are one of the most  
4 common veterinary species to exhibit coronary artery anomalies, with examples of such abnormalities  
5 including coronary-to-pulmonary artery fistulae and anomalous origin of the left coronary artery from the  
6 pulmonary trunk. Consequently, their usage as animal models to mimic the human coronary system merits  
7 careful scrutiny before findings can be extrapolated.<sup>38,39</sup>

## 9 **Suitability for use as animal clinical models in Type-A aortic** 10 **dissection research**

11 Having explored the anatomical differences between ovine, bovine and the porcine species, the anatomic  
12 feasibility of using these as animal models to better our understanding of Type-A AD treatment options shall  
13 now be explored.

14  
15 As aforementioned, Type-A ADs involve the ascending aorta, making this aspect of the model's anatomy  
16 significantly important. Bovine aortic anatomy is particularly unhelpful for this pathology therefore, given the  
17 marked differences compared to their human counterparts, as elucidated previously.<sup>28</sup> Indeed, the 'bovine  
18 aortic arch effect' is an epidemiological term used to highlight the linkage between ascending and thoracic  
19 aortic dilatation as a result of the aortic arch anatomy within bovines, further exemplifying their unsuitability as  
20 an animal model in this context.<sup>40</sup>

21  
22 Between the ovine and porcine species, it appears that each share some features with that of humans, whilst  
23 also exhibiting some differences that impact their usage as animal models. For instance, whilst ovines  
24 maintain a uniform aortic diameter similar to their human counterparts, their short immobile aorta could pose a  
25 challenge to graft repair within animal models.<sup>27</sup> Conversely, in spite of the larger aorta within pigs, the aortic  
26 diameter being nearly a fifth lesser than that of their human counterparts could also affect reproducibility of  
27 findings to the latter. Consequently, it is difficult to assess which of ovine or porcine models is better for  
28 modelling Type-A ADs, at least from the ascending aortic anatomy point-of-view.

29  
30 The aortic valvular anatomy also holds certain significance when choosing an appropriate animal model,  
31 particularly with AD tears extending proximally into the aortic root.<sup>41</sup> As aforementioned, variations in leaflet  
32 thickness are of importance, as the heavy pressure loads exerted during clinical usage with can affect the  
33 structural stability of the animal model. Consequently, species with relatively thinner valvular commissures,  
34 such as in ovines, must be handled with due care, and it is for this reason that porcine models are preferred to  
35 their counterparts.

36  
37 Finally, the coronary vasculature of the aforementioned animal models also has particular relevance to the  
38 pathology of Type-A ADs, especially with tears arising in the aortic root, or even with any more distal tears  
39 causing dissections in the proximal sinotubular junction, both of which would affect the coronary supplies, and  
40 thus consequently cause ischemia of the cardiac musculature. Given that bovine species exhibit the most  
41 coronary artery anomalies, their usage as an animal model in better understanding the various treatment

1 options for Type-A ADs is hence not justified, given that these findings would not necessarily accurately  
2 represent what we might see within humans.[38,39](#)

3  
4 Between porcine and ovine species however, the coronary vasculature is similar to that of humans. However,  
5 as aforementioned, much more research has been performed on the coronary arterial supply of pigs, with little  
6 to no comparative research being performed on their ovine counterparts, and as such, the former takes  
7 current precedence when selecting an animal model for Type-A AD research.

8

## 9 **Limitations of this review & insights on future research**

10 The comparison of ovine, porcine and bovine cardiac anatomy, and their usage as animal models will  
11 undoubtedly provide important new insights in new endovascular treatment options within the Type-A AD  
12 paradigm. However, as explored in this paper, several limitations exist, with a prominent example being the  
13 lack of literature in anatomical differences amongst each of these three species. Firstly, there is a lack of  
14 information in the microscopic anatomical differences between species' cardiac anatomy, such as the  
15 anatomical variances in the layers of the aorta amongst porcine, ovine and bovine species. Additionally,  
16 although considerable literature describes either very general or very specific aspects of mammalian cardiac  
17 anatomy, little quantitative, truly comparative research has been done. These ties in to our final limitation,  
18 which is the nature of this review itself. As a narrative review, whilst it provides information about the current  
19 stage of research and addresses future directions and possible clinical applications, it has a limited  
20 comprehensive results analysis. Potentially, a systematic review might yield more comprehensive data as well  
21 as identify any bias or random errors. In the long term, the authors encourage researchers currently using  
22 animal models of cardiovascular disease to publish their findings and add to the literature to allow such  
23 translation to human interventions.

## 24 **Conclusion**

25 The introduction of intravascular stent-grafts as a surgical treatment option for Type-A ADs represents one of  
26 the most successful innovations in cardiothoracic surgery within the last few decades. However, lingering high  
27 numbers of patient mortality rates in spite of surgical intervention highlights the accelerated need for our better  
28 understanding of novel treatment options for this disease, explicating the necessity of developing an  
29 appropriate animal clinical model. From a strictly anatomical standpoint, bovine species do not meet this  
30 need, given the significant variations in aortic arch anatomy, the lack of literature on aortic valvular anatomy  
31 and finally the significant variation in coronary artery anatomy. However, both porcine and ovine species  
32 appear to be suitable options to be used as animal models for proximal aortic endovascular treatment, with  
33 the former possessing a slight advantage given similarities in coronary artery and aortic valve anatomy to their  
34 human analogues. The identification of appropriate animal models will provide knowledge for further insight  
35 into the available endovascular treatment options into Type-A ADs, and consequently needs to be hastened.

1 **REFERENCES.**

- 2 1. Cesarovic N, Lipiski M, Falk V, Emmert M. Animals in cardiovascular research: Clinical relevance and  
3 translational limitations of animal models in cardiovascular medicine. *EHJ*. 2020; 41(2):200-3.
- 4 2. Criado F. Aortic Dissection: A 250-Year Perspective. *Tex Heart Inst J*. 2011; 8(6):694-700.
- 5 3. Fujimura N, Kawaguchi S, Obara H, Yoshitake A, Inoue M, Otsubo S et al. Anatomic Feasibility of  
6 Next-Generation Stent Grafts for the Management of Type A Aortic Dissection in Japanese Patients.  
7 *Circ J*. 2017; 81:1388–94.
- 8 4. Chiu P, Miller DC. Evolution of surgical therapy for Stanford acute type A aortic dissection. *Ann*  
9 *Cardiothorac Surg*. 2016; 5(4):275-95.
- 10 5. Scholl F, Coady M, Davies R. Interval or Permanent Nonoperative Management of Acute Type A  
11 Aortic Dissection. *JAMA Surgery*. 1999; 134(4):402-6.
- 12 6. Auer J, Berent R, Eber B. Aortic Dissection: Incidence, Natural History and Impact of Surgery. *Journal*  
13 *of Clinical and Basic Cardiology*. 2000; 3(3):151-4.
- 14 7. Fann JI, Smith JA, Miller DC, et al. Surgical management of aortic dissection during a 30-year period.  
15 *Circulation* 1995; 92(2):113.
- 16 8. Becker H, Jauch K. *Vascular Surgery*. 1st Edition. Berlin: Springer-Verlag; 1989. p. 349-60
- 17 9. Shah A, Khoynezhad A. Thoracic endovascular repair for acute type A aortic dissection: operative  
18 technique. *Ann Cardiothorac Surg*. 2016; 5(4):389-96.
- 19 10. Kreibich M, Rylski B, Kondov S, Morlock J, Scheumann J, Kari F et al. Endovascular treatment of  
20 acute Type A aortic dissection—the Endo Bentall approach. *J Vis Surg*. 2018; 1(4):69.
- 21 11. Heilmann C, Stahl R, Schneider C, Sukhodolya T, Siepe M, Olschewski M et al. Wound complications  
22 after median sternotomy: a single-centre study. *Interact Cardiovasc Thorac Surg*. 2013; 16(5):643-8.
- 23 12. Luciani G, Lucchese G. Minimal-access median sternotomy for aortic valve replacement. *J Thorac*  
24 *Dis*. 2013; 5(Suppl 6): S650–3.
- 25 13. Sarkar M, Prabhu V. Basics of cardiopulmonary bypass. *Indian J Anaesth*. 2017;61(9):760–7.
- 26 14. Zanotti G, Reece TB, Aftab M. Aortic Arch Pathology: Surgical Options for the Aortic Arch  
27 Replacement. *Cardiol Clin*. 2017; 35(3):367-85.
- 28 15. Nordon IM, Hinchliffe RJ, Morgan R, Loftus IM, Jahangiri M, Thompson MM. Progress in  
29 endovascular management of type A dissection. *Eur J Vasc Endovasc Surg*. 2012; 44(4):406-10.
- 30 16. Kreibich M, Soekeland T, Beyersdorf F, Bavaria J, Schröfel H, Czerny M et al. Anatomic feasibility of  
31 an endovascular valve-carrying conduit for the treatment of type A aortic dissection. *J Thorac*  
32 *Cardiovasc Surg*. 2019; 157(1):26-34.e1.
- 33 17. Harky A, Al-Adhami A. Stenting in type A aortic dissection: fantasy or reality? *J Vis Surg*. 2018;  
34 4(161):1-3.
- 35 18. Mangialardi N, Serrao E, Ronchey S, Kasemi H, Orico M. Endovascular Treatment of Type A  
36 Dissections. *Endovascular Today*. 2013 Nov. Available from: [https://evtoday.com/articles/2013-](https://evtoday.com/articles/2013-nov/endovascular-treatment-of-type-a-dissections)  
37 [nov/endovascular-treatment-of-type-a-dissections](https://evtoday.com/articles/2013-nov/endovascular-treatment-of-type-a-dissections)
- 38 19. University of Minnesota. Comparative Anatomy of the Valves. Available from:  
39 <http://www.vhlab.umn.edu/atlas/comparative-anatomy-tutorial/external-anatomy.shtml>. Last updated  
40 [Jan 14,2019]; cited [Jan 20,2020].
- 41 20. Sands M, Rittenhouse E, Mohri H, Merendino K. An Anatomical Comparison of Human, Pig, Calf, and  
42 Sheep Aortic Valves. *Ann Thorac Surg*. 1969; 8(5):407-14.
- 43 21. University of Minnesota. Comparative Anatomy of the Valves. Available from:  
44 <http://www.vhlab.umn.edu/atlas/comparative-anatomy-tutorial/valves.shtml>. Last updated Jan  
45 14,2019; cited Jan 20,2020.
- 46 22. Wang C, Lachat M, Regar E, von Segesser L, Maisano F, Ferrari E. Suitability of the porcine aortic  
47 model for transcatheter aortic root repair. *Interact Cardiovasc Thorac Surg*. 2017; 26(6):1002-8.
- 48 23. Tao L, Xianhao B, Yuxi Z, Ziwen L, Ziyi X, Zhaoxiang Z et al. Thoracic aortic computed tomography  
49 angiography in porcine: establishment of a baseline for endovascular evaluation of the ascending  
50 aorta. *Interact Cardiovasc Thorac Surg*. 2020;31(2):248-53
- 51 24. Khan S, Islam M. Studies on the Prospect of Bioprostheses by Bovine Aortic Valve for Human Use.  
52 *Bangladesh Med Res Counc Bull*. 1991; 17(2):75-80

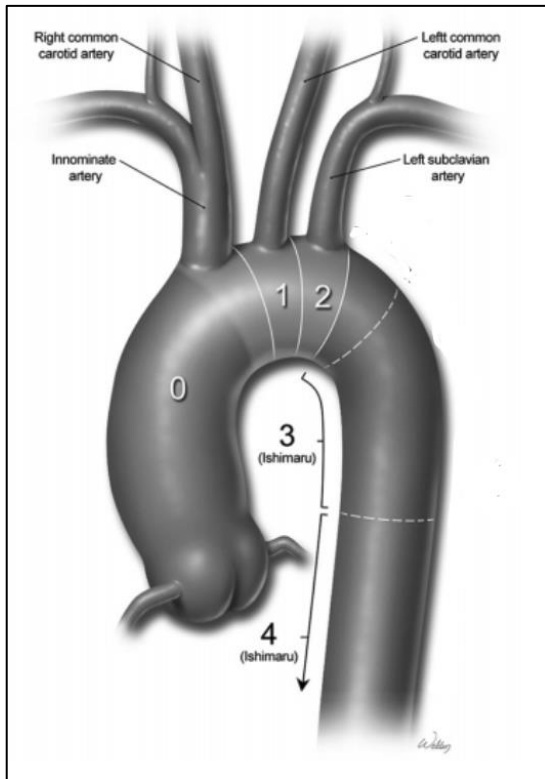
- 1 25. Hyun Joh J, Ahn H, Park H. Reference Diameters of the Abdominal Aorta and Iliac Arteries in the
- 2 Korean Population. *Yonsei Med J.* 2013; 54(1):48-54.
- 3 26. Jonker F, Mojibian H, Schlösser F, Botta D, Indes J, Moll F et al. The Impact of Hypovolaemic Shock
- 4 on the Aortic Diameter in a Porcine Model. *Eur J Vasc Endovasc Surg.* 2010; 40(1):564-71.
- 5 27. DiVincenti L, Westcott R, Lee C. Sheep (*Ovis aries*) as a Model for Cardiovascular Surgery and
- 6 Management before, during, and after Cardiopulmonary Bypass *J Am Assoc Lab Anim Sci.* 2014;
- 7 53(5):439-48.
- 8 28. Dumfarth J, Chou A, Ziganshin B, Bhandari R, Peterss S, Tranquilli M et al. Atypical aortic arch
- 9 branching variants: A novel marker for thoracic aortic disease. *J Thorac Cardiovasc Surg.* 2015;
- 10 149(6):1586-92.
- 11 29. Layton K, Kallmes D, Cloft H, Lindell E, Cox V. Bovine Aortic Arch Variant in Humans: Clarification of
- 12 a Common Misnomer. *AJNR Am J Neuroradiol.* 2006; 27(7):1541-2.
- 13 30. Torad F, Amer M, Shamaa A, Elsherpieny E. Echocardiographic measurements and indices in normal
- 14 adult buffalo (*Bubalus bubalis*). *Journal of Applied Animal Research.* 2016;45(1):336-41.
- 15 31. Devereux R, Simone G, Arnett D, Best L, Boerwinkle E, Howard B et al. Normal Limits in Relation to
- 16 Age, Body Size and Gender of Two- Dimensional Echocardiographic Aortic Root Dimensions in
- 17 Persons  $\geq 15$  Years of Age. *Am J Cardiol.* 2012; 110(8):1189-94.
- 18 32. Braun U, Schweizer T. Determination of Heart Dimensions in Cattle via 2-D-mode Echocardiography.
- 19 *Berl Munch Tierarztl Wochenschr.* 2001;114(2):46-50.
- 20 33. Sahni D, Kaur G, Jit H, Jit I. Anatomy & Distribution of Coronary Arteries in Pig in Comparison With
- 21 Man. *Indian J Med Res.* 2008;127(6):564-70/
- 22 34. Weaver M, Pantely G, Bristow J, Ladley H. A Quantitative Study of the Anatomy and Distribution of
- 23 Coronary Arteries in Porcine in Comparison With Other Animals and Man. *Cardiovasc Res.*
- 24 1986;20(12):907-17.
- 25 35. Gómez F, Ballesteros L. Evaluation of coronary dominance in pigs; a comparative study with findings
- 26 in human hearts. *Arq. Bras. Med. Vet. Zootec.* 2015;67(3):783-9.
- 27 36. Frink R, Merrick B. The Sheep Heart: Coronary and Conduction System Anatomy With Special
- 28 Reference to the Presence of an Os Cordis. *Anat Rec.* 1974;179(2):189-200.
- 29 37. Scansen B. Coronary Artery Anomalies in Animals. *Vet. Sci.* 2017;4(2):20.
- 30 38. Barszcz K, Polgaj M, Klećkowska-Nawrot J, Goździewska-Harłajczuk K, Olbrych K, Czopowicz M.
- 31 Morphometry and topography of the coronary ostia in the European bison. *Folia Morphol.*
- 32 2019;79(1):105-12.
- 33 39. Gómez F, Cortés L, Ballesteros L. Morphological characterisation of the coronary arteries in African
- 34 sheep (*Ovis orientalis*). Differential analysis with those of humans and other animal species. *Folia*
- 35 *Morphol.* 2018;78(1):63-70.
- 36 40. Pham T, Martin C, Elefteriades J, Sun W. Biomechanical characterisation of ascending aortic
- 37 aneurysm with concomitant bicuspid aortic valve and bovine aortic arch. *Acta Biomater.*
- 38 2013 ;9(8):7927-36.
- 39 41. Ho S. Structure and anatomy of the aortic root. *Eur J Echocardiogr.* 2009;10(1):3-10.
- 40

1 **FIGURES AND TABLES.**

2

3 **Figure 1.** Ishimaru Classification of Various Landing Zones of Proximal Aorta for Endovascular Arch Repair

4



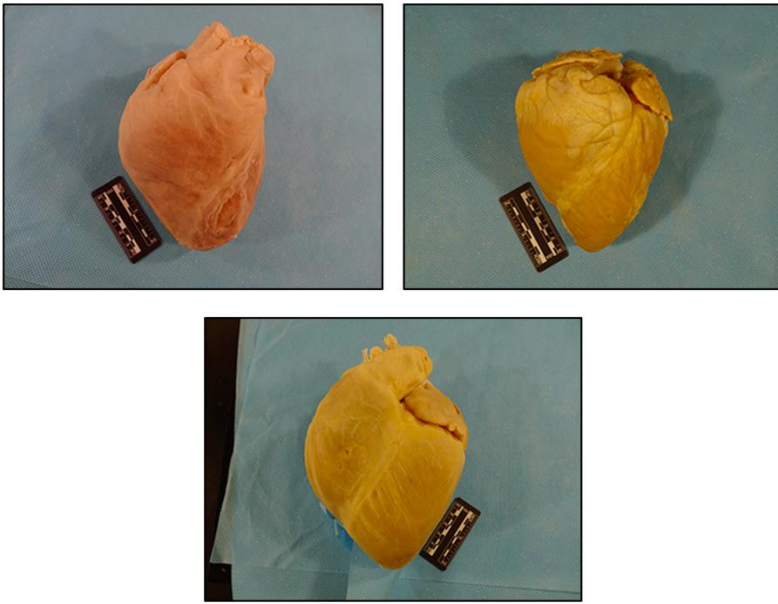
5

6 **Reference:** Zanotti G, Reece TB, Aftab M. Aortic Arch Pathology: Surgical Options for the Aortic Arch

7 Replacement. *Cardiol Clin.* 2017; 35(3):367-85.



1 **Figure 2.** Plastinated Human (upper left), Ovine (upper right) and Porcine (bottom) Hearts



2

3 **Reference:** University of Minnesota. Comparative Anatomy of the Valves. Available from:

4 <http://www.vhlab.umn.edu/atlas/comparative-anatomy-tutorial/valves.shtml>. Last updated [Jan 14,2019]; cited

5 [Jan 20,2020]

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1 **Table 1.** Mean Dimensions and Standard Deviations of Aortic Valve Measurement

Measurement		Human	Porcine	Bovine	Ovine
Annulus diameter of aortic valve (obturator diameter)		26.4 ± 3.15 <sup>20</sup>	26.6 ± 1.84 <sup>20</sup>	33.7 ± 2.74 <sup>20</sup>	25.8 ± 1.29 <sup>20</sup>
Leaflet depth	Non-coronary cusp	9.1 ± 1.66 <sup>20</sup>	8.9 ± 1.46 <sup>20</sup>	9.2 ± 1.58 <sup>20</sup>	7.4 ± 1.36 <sup>20</sup>
	Right coronary cusp	9.8 ± 2.21 <sup>20</sup>	10.2 ± 1.45 <sup>20</sup>	9.9 ± 1.21 <sup>20</sup>	7.6 ± 1.26 <sup>20</sup>
	Left coronary cusp	9.3 ± 1.24 <sup>20</sup>	8.6 ± 1.56 <sup>20</sup>	9.9 ± 0.96 <sup>20</sup>	7.8 ± 1.77 <sup>20</sup>
Valvular commissure height	Non-coronary cusp	18.5 ± 1.96 <sup>20</sup>	14.9 ± 1.84 <sup>20</sup>	19.5 ± 1.92 <sup>20</sup>	13.7 ± 1.52 <sup>20</sup>
	Right coronary cusp	17.5 ± 2.95 <sup>20</sup>	17.3 ± 2.28 <sup>20</sup>	19.4 ± 1.57 <sup>20</sup>	13.4 ± 1.75 <sup>20</sup>
	Left coronary cusp	17.3 ± 2.61 <sup>20</sup>	16.3 ± 2.00 <sup>20</sup>	19.1 ± 2.53 <sup>20</sup>	13.9 ± 1.30 <sup>20</sup>

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3

*NB. All measurements in the table above are in millimetres.*

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1 **Table 2.** Dimensions of the Aorta

Measurement	Human	Porcine	Bovine	Ovine
Aortic annulus diameter	23.0 ± 2.5 <sup>21</sup>	20.0 ± 1.2 <sup>21</sup>	48.0 ± 0.92 <sup>24</sup>	Not documented in adults
Thoracic aortic diameter at sinotubular junction	27.2 ± 3.0 <sup>21</sup>	20.0 ± 0.9 <sup>21</sup>	Not documented in adults	Not documented in adults
Abdominal aorta diameter (measured at level of superior mesenteric artery)	22.0 ± 0.3 <sup>25</sup>	10.4 <sup>21</sup>	Not documented in adults	Not documented in adults

2

3 *NB. All measurements in the table above are in millimetres. Standard deviations for abdominal aortic*  
 4 *dimensions in pigs were not documented.*

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1 **Table 3.** Dimensions of the Coronary Vasculature

Measurement	Human	Porcine	Bovine	Ovine
Left coronary ostia diameter	4.8 ± 0.5 <a href="#">21</a>	5 ± 0.5 <a href="#">21</a>	7.1 ± 1.7 <a href="#">38</a>	5.38 ± 1.59 <a href="#">39</a>
Right coronary ostia diameter	3.7 ± 0.9 <a href="#">21</a>	4.7 ± 0.5 <a href="#">21</a>	5.3 ± 1.4 <a href="#">38</a>	1.75 ± 0.44 <a href="#">39</a>
Coronary collateralization	Limited	Limited	Anomalous	Limited

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3

*NB. All measurements in the table above are in millimetres*

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